The effectiveness of plant essential oils on the *in vitro* growth of postharvest phytopathogenic fungi



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Background and objectives

Plant extracts, such as essential oils (Eos) [1], which have long been used in traditional preparations, are currently adopted by the industrial manufactures for modern formulations of consumer products. They are found in perfumery and cosmetology as well as in food and pharmaceutical industries. In agriculture, it was reported that Eos could be an interesting alternative to chemical fungicides and could be used as biofungicides in postharvest biological control. The present work was therefore undertaken to study the effect of thirty species of EOs on the *in vitro* growth of *Penicillium italicum*, *Penicillium digitatum* infecting citrus (fig.1 & fig.2) and *Colletotrichum musea* infecting bananas (fig.3).



Figure 1 : Blue mold

(P. italicum)

*Obtained from Pranarôm International S A



Figure 2 : Green mold (P. digitatum) Figure 3 : Anthracnose

(C. musea)

x10 x3 x1 x4 x8 x6 x6 x9

X1 X7 X3

Plate 2

Main constituents of essential oils (terpenes and aromatics compounds)	Essential oils*	Origin	Part of the plant distilled				
	1	Italia	Zest				
Non-oxygenated compounds	2	Bearl	Zest				
	3	Ceylon	Fruit				
	11	Beam	Rhizonse				
	4	Egypt	Flowering top				
	5	France	Arial part				
Alcohoh	6	India	Arial part				
	13	Morocce	Flowering top				
	14	Australia	Leaf or leaves				
	16	France	Arial part				
	1	Guatemala	Arial part				
Aldehydes	17	Ceylen	Bark				
	18	Ceylen	Leaf				
Ketones	8	Maracce	Arial part				
	12	USA	Arial part				
	18	France	Flowering top				
	19	Madagascar	Bed				
	20	India	Fruit				
Phenoh	21	Ceska	Arial part				
	22	Spain	Flowering top				
	23	Maracco	Flowering top				
	24	Morocco	Flowering top				
	25	France	Flowering top				
Esters	9	France	Flowering top				
	10	hingy	Fruit				
	26	Vietnam	Flowering top				
Ether oxides	27	Chine	Leaf				
	28	France	Arial part				
	29	Madagascar	Leaf				
		the second se	the second se				

Table 1: Essential oils

X10	хз	XI	X 4			т	π	17	в	n	75	n	Π	ъ	п	75	г	
xı	X.6	XS	33				19	T4	TS	T10	Té	19	T4	TS	TIO	T6		
xı	X7	X3	x 2		x													x.
X5	X5	X9	X4					X10	X3	XI	X4	X10	X3	xı	X4			
X2 X8	X8	X7	X10					XS	X6	X6	X9	33	XS	X6	X9			
						x.		xı	X7	X3	X2	XI	X7	х3	X2		x.	
TI	π	в	n	TS				xs	X5	X9	X4	xs	X5	X9	<u>X</u> 4			
T 9	T4	TS	TIO	T6	т			X2	XS	X7	хю	X2	XB	X7	X10			г

Plate 1

h EO T: Solution without microorganism a



Material and methods

Essential oils were selected according to intrinsic (yield, phytotoxicity) and extrinsic (availability, cost, popularity) criteria [2], and were classified according to their main constituents (table 1). Their effect on the growth of three pathogens was evaluated with decreasing concentrations of application, using a microdilution method on a 96-well microplate Elisa (figure 4). Only essential oils inhibiting more than 70 % of the growth of all the fungi were retained for the following test. For each pathogen, growth was followed by recording each day the optical density (OD) at 490 nm of a solution of 200 μ l containing: diluted orange juice (0,03 v/v); 10⁴ conidia/ml; essential oil (1000, 500 and 100 ppm).

The experimental design is shown in figure 4. For each pathogen, two independent experiments were performed with 8 replicates per essential oil. The mean % of inhibition of essential oils was calculated as follows:

 $\left[OD\left(Xn\right)-OD\left(Tn\right)\right]-\left[OD\left(X'\right)-OD\left(T'\right)\right]$

Mean % inhibition =

[OD (X') – OD (T')

[OD (Xn) – OD (Tn)] = OD of the pathogen in presence of EO [OD (X') – OD (T')] = OD of the pathogen in absence of EO

Results

Only recorded data of the growth period 168-192 h are shown in Figure 5.
Whatever the essential oil tested and whatever the concentration (except EO 20 tested at 500 ppm), there was a growth inhibition regardless of the species.
In general, the three pathogens present different sensitivity levels towards the EOs. *C. musea* seems to be more sensitive than the tested *Penicillium* species.
All pathogens appear to be more sensitive to EOs composed of alcohol, aldehyde or phenol.

Among the 30 essential oils tested, 12 were effective against all the fungi at 1000 ppm and 3 at 100 ppm.

Conclusion

The present preliminary work shows that essential oils are able to partially or totally inhibit the growth of three different post-harvest fungal pathogens. This is a first encouraging study for the development of biofungicides based on essential oils as an alternative to chemical fungicides.

References

[1] Ernest Guenther. The essential oils - vol 1: History – Origin In Plants – Production – Analysis. Read Book (2007) 452 Pages.

[2] Pranarôm International S.A, expert en aromathérapie scientifique et médicale, Tarif export 2008.